# **Original Article**

# *GC* gene polymorphisms found with type 2 diabetes and low vitamin D status among rural Chinese in Henan province

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Background and Objectives: Accumulating evidence suggests that vitamin D may be involved in the pathogenesis of type 2 diabetes (T2D). Group-specific component (GC) gene is the most important transporter of vitamin D and plays a regulatory role in vitamin D metabolism. We aimed to evaluate the association of GC gene polymorphisms with T2D susceptibility and vitamin D status in the Chinese rural population. Methods and Study Design: A total of 1372 subjects were eligible in this cross-sectional study. Three SNPs of the GC gene (rs7041, rs4588, and rs2282679) were genotyped by TaqMan probe assays. Logistic regression and Kruskal-Wallis oneway analysis were performed to determine the possible risk genotype for T2D and vitamin D metabolite concentrations, respectively. Results: The serum 25-hydroxyvitamin D<sub>3</sub> [25(OH)D<sub>3</sub>] and vitamin D binding protein (DBP) concentrations were significantly lower in the T2D group than the non-T2D group. GG genotype carriers of rs7041 (T>G) were more likely to have T2D compared with AA carriers (OR=2.00, 95% CI: 1.19-3.37). Variants of rs4588 (C>A) and rs2282679 (A>C) were associated with a lower risk of T2D under the dominant inheritance model (OR=0.65, 95% CI: 0.48-0.88; OR=0.66, 95% CI: 0.49-0.90, respectively). We further found that non-T2D subjects with the AA genotype of rs4588 had significantly higher 25(OH)D<sub>3</sub> concentrations than the CC genotype (p=0.022). In contrast, the T2D cases with the CC genotype of rs2282679 had lower DBP concentrations compared to the AA genotype (p=0.020). Conclusions: Our study indicates a potential role for GC gene polymorphisms in T2D susceptibility and vitamin D metabolite concentrations in the Chinese rural population.

Key Words: GC gene, polymorphism, type 2 diabetes, vitamin D binding protein, 25(OH)D<sub>3</sub>

#### INTRODUCTION

Type 2 diabetes (T2D) is a complex metabolic disease that is characterized by insulin resistance, defective insulin secretion, or both of them.<sup>1</sup> The 9th edition of the International Diabetes Federation Diabetes Atlas predicts that the number of diabetic patients will increase to 578 million by 2030 and 700 million by 2045.<sup>2</sup> T2D can lead to serious complications such as cardiovascular disease, stroke, kidney failure, and retinopathy.<sup>3</sup> T2D is a multifaceted disorder with several risk factors, including genetics, diet, and personal behaviors, of which genetics plays a critical role with an estimated heritability of 40% to 80%.<sup>4</sup>

As the main circulating vitamin D metabolites, serum 25-hydroxyvitamin D3  $[25(OH)D_3]$  is a clinical biomarker for judging the status of vitamin D. Vitamin D binding protein (DBP) is a protein encoded by GC gene, which is converted into various forms by combining with vitamin D and transported to various organs of the body.<sup>5</sup> GC is located on chromosome 4q13 and has 13 exons, encoding 474 amino acids. There are three common single nucleotide polymorphisms (SNPs) of the GC gene, which are rs7041 (T>G) and rs4588 (C>A) located in exon 11 and rs2282679 (A>C) located in intron 12.<sup>6</sup> DBP concentrations are susceptible to GC gene polymorphism,

which affects the affinity of vitamin D and its metabolites, resulting in changes in the concentration of 25(OH)D<sub>3</sub>.<sup>7</sup> Studies have demonstrated that the GC gene was involved in circulating 25(OH)D concentration.<sup>8,9</sup>

Due to the biological function of the GC gene, numerous researchers tend to explore diseases related to vitamin D concentrations with the polymorphism of the GC gene, such as cancer, coronary artery disease, Parkinson's disease, and Alzheimer's disease.<sup>10-12</sup> In addition, related studies have also been conducted to investigate the relationship between GC SNPs and T2D susceptibility, but the existing findings are inconsistent. Wang, et al. and Rahman, et al. demonstrated that GC SNPs were associated with an increased risk of T2D in Asians and Bangladesh, respectively.<sup>13,14</sup> Whereas Ye et al and Malecki et al

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did not find the evidence for an association between GC gene SNPs and T2D susceptibility in French Caucasians and Poland.<sup>15,16</sup>

Thus, the objective of the current study was to investigate the associations of (1) GC SNPs and T2D risk in a cross-sectional study; (2) the vitamin D-related metabolite concentrations and T2D risk; (3) serum vitamin Drelated metabolite concentrations and GC SNPs.

# METHODS

## Subjects

The subjects of this cross-sectional study were recruited by the method of cluster random sampling. Two towns, Wuzhi in Jiaozuo City and Houzhai in Zhengzhou City, were randomly selected from rural areas of Henan Province. Ultimately, a total of 1570 subjects were willing to participate and complete the examination from July to August 2013 and July to August 2015. The present analysis excluded individuals younger than 18 years of age (n=102), failing in DNA extraction (n=24), or vitamin D extreme values (n=72). Finally, we obtained a sample of 1372 participants. The detailed flow chart of the research object is shown in Figure 1. We collected data with basic information such as age, gender, physical activity, history of chronic diseases, culture, and vitamin D supplement intake during face-to-face interviews. They then completed a physical examination (height, weight, waist circumference, and hip circumference), fasting blood glucose, and blood lipid tests. According to the definition and standards of the World Health Organization (WHO),<sup>17</sup> T2D was defined as fasting plasma glucose  $\geq$ 7.0 mmol/L and/or taking hypoglycemic drugs and/or having a selfreported history of T2D, we eventually identified 231 cases of T2D patients.

This study was approved by the Zhengzhou University Life Science Ethics Committee (Code: [2015] MEC (S128)). Written informed consent was obtained from all participants.

# **Biochemical measurements**

Fast blood glucose and lipid were determined by the au-

tomatic biochemical analyzer (KHB360, Shanghai, China). Serum 25(OH)D<sub>3</sub> and DBP concentrations were detected by ELISA kits (Sangon Biotech, Shanghai, PR China). The absorbance of the microplate reader at 450 nm was measured. According to the Institute of Medicine (IOM),<sup>18</sup> we classified serum 25(OH)D<sub>3</sub> into three groups: vitamin D deficiency (VDD) (<20 ng/mL), vitamin D insufficient (VDI) (20-30 ng/mL), vitamin D sufficient (VDS) ( $\geq$ 30 ng/mL).

#### Genotyping

Genomic DNA was extracted from peripheral blood according to the standard procedures (DNA blood kit, Biotech, Beijing, China). Based on the HapMap website and previous research, three SNPs of the GC gene (rs7041, rs4588, and rs2282679) were identified for further analysis. Genotyping was completed by TaqMan probe assays and employing an Applied Biosystems (ABI, 7500 FAST Real-time PCR system, Foster City, USA) platform. For genotyping quality control, we selected 10% random samples for duplicated analysis, and the concordance rate was 99.5%.

#### Sample size and statistical analyses

We used the previous study of GC variants and T2D reported by Fawzy<sup>19</sup> to obtain a rough estimate of the odds ratio (OR=2.50) and considered the minor allele frequencies (MAF) of rs2282679 as the prevalence of risk factor (p=0.178). Based on this, we estimated a required sample size of 124 to detect an effect of similar magnitude at an  $\alpha$  level of 0.05 with 90% statistical power.

Statistical analyses were performed using IBM SPSS 21.0 (SPSS, Chicago, IL, USA). Continuous variables conforming to normality were compared by Student's t-test for two groups, which were represented by means  $\pm$  standard deviations (SD). Otherwise, Wilcoxon rank-sum test was used, and the data were expressed as medians (interquartile ranges). Categorical variables were assessed using the chi-square test. We used the goodness-of-fit  $\chi^2$  test to examine Hardy–Weinberg equilibrium. Linkage



Figure 1. Flow chart of the research object source.

disequilibrium was determined using Haploview. Logistic regression models were performed to evaluate the relationship between the three SNPs of the GC gene and the risk of T2D. The results were corrected for multiple comparisons using a false discovery rate (FDR). Kruskal-Wallis one-way analysis was performed to estimate the association between serum 25(OH)D3 and DBP concentrations and GC SNPs. Multivariate ORs and 95% confidence intervals (CI) for T2D according to the divided serum vitamin D metabolite quartiles were calculated using a logistic regression while adjusting for age, gender, married status, smoking status, drinking status, high-fat intake, physical activity, and family history of diabetes. We also conducted subgroup analysis based on age, gender, family history of diabetes, hypertension or not, and vitamin D status to further identify the robustness of our results. A two-sided p-value <0.05 was considered to be statistically significant.

### RESULTS

#### Characteristics of the subjects

A total of 231 individuals with T2D and 1141 non-T2D fulfilled the criteria and were included in the analysis. The T2D group showed higher concentrations of total cholesterol (TC), triglyceride (TG), and low-density lipoprotein cholesterol (LDL-C) than the non-T2D group. Inversely, the high-density lipoprotein (HDL-C), serum 25(OH)D<sub>3</sub> and DBP concentrations of T2D patients were lower than that of the non-T2D group (p<0.05). There was a significant difference in the distribution of vitamin D status between the T2D and non-T2D groups (p=0.001). The percentage of T2D patients with VDD and VDI was 74.0%. No significant difference was found in the family history of diabetes between the two groups (p=0.403).

Table 1. Baseline characteristics of study participants

Clinical characteristics of the T2D and the non-T2D group are presented in Table 1.

#### Association between GC polymorphisms and T2D

All genotype frequencies of the two groups were in Hardy–Weinberg equilibrium (p>0.05). The results of linkage disequilibrium between the three SNPs are shown in Supplementary Figure 1. Table 2 summarizes the genotype distribution of three SNPs between the T2D and non-T2D groups. The three SNPs (rs7041/rs4588/rs2282679) were all significantly different in the distribution of the two groups (p=0.007, 0.020, and 0.022, respectively). Moreover, rs4588 (C>A) and rs2282679 (A>C) were associated with a lower risk of T2D under the dominant inheritance model (OR=0.65, 95% CI: 0.48-0.88; OR=0.66, 95% CI: 0.49-0.90). Notably, for the rs7041 (T>G) variant, carriers of the GG genotype were more likely to suffer from T2D compared with those carrying the TT genotype (OR=2.00, 95% CI: 1.19-3.37).

To further explore the effect of other risk factors on the association of GC gene SNPs with T2D risk, subgroup analyses were performed based on age, gender, family history of diabetes, hypertension or not, and vitamin D status (Table 3). Overall, the associations of SNPs and T2D were consistent in most of the subgroups. In addition, rs7041 polymorphism was associated with a higher risk of T2D in subjects with a family history of diabetes and without hypertension. It should be noted that rs4588 and rs2282679 were associated with a lower risk of T2D in females, subjects with age <60 years and no family history of diabetes, under the dominant inheritance model.

#### Vitamin D metabolite concentrations

We analyzed the distribution of vitamin D metabolites in the three SNP genotypes in T2D and non-T2D groups

Characteristics	T2D (n=231)	Non-T2D (n=1141)	Statistics	p value
Gender			6.33	$0.012^{\dagger}$
Male	96 (41.7%)	580 (50.8%)		
Female	134 (58.3%)	561 (49.2%)		
Age (years)			66.6	$<\!\!0.001^{\dagger}$
<u>≤</u> 40	18 (7.80%)	354 (31.0%)		
40-60	90 (39.0%)	437 (38.3%)		
≥60	123 (53.3%)	350 (30.7%)		
TC (mmol/L)	4.61 (3.95, 5.51)	4.34 (3.68, 5.06)	12.4	0.001‡
TG (mmol/L)	1.69 (1.05, 2.67)	1.17 (0.76, 1.92)	40.5	<0.001‡
HDL-C (mmol/L)	1.27 (1.10, 1.47)	1.34 (1.15, 1.55)	8.88	0.003‡
LDL-C (mmol/L)	2.49 (1.97, 3.09)	2.32 (1.85, 2.87)	5.09	0.024‡
25(OH)D <sub>3</sub> (ng/mL)	18.4 (14.9, 32.4)	21.1 (15.2, 50.4)	7.01	$0.008^{\ddagger}$
DBP (ng/mL)	124 (99.2, 239)	155 (103, 399)	11.6	0.001‡
Family history of diabetes			1.82	$0.403^{\dagger}$
Yes	53 (22.9%)	264 (23.1%)		
No	174 (75.3%)	886 (76.1%)		
Vitamin D status			13.6	$0.001^{+}$
VDD	129 (55.8%)	540 (47.3%)		
VDI	42 (18.2%)	160 (14.0%)		
VDS	60 (26.0%)	441 (38.7%)		

DBP: vitamin D binding protein; VDD: Vitamin D deficiency; VDI: vitamin D insufficient; VDS: vitamin D sufficient.

Data are expressed as the frequency and percentage (%) for categorical variables or medians (interquartile ranges) for continuous variables with non-normal distributions.

<sup>†</sup>Chi-square test.

<sup>‡</sup>Wilcoxon rank sum test.

SNPs	T2D	Non-T2D	$\chi^2$	$p^{\dagger}$	OR (95% CI)‡	$p^{\S}$	$p_{-FDR}$	$p_{-HWE}$
rs7041			9.94	0.007				0.162
TT	114 (49.4%)	638 (55.9%)			Reference			
TG	90 (39.0%)	433 (38.0%)			1.12 (0.82, 1.55)	0.472	0.472	
GG	27 (11.7%)	70 (6.10%)			2.00 (1.19, 3.37)	0.010	0.030	
TG+GG					1.25 (0.93, 1.68)	0.147	0.221	
rs4588			7.78	0.020				0.763
CC	126 (54.6%)	510 (44.7%)			Reference			
CA	88 (38.1%)	514 (45.1%)			0.70 (0.51, 0.96)	0.028	0.028	
AA	17 (7.40%)	117 (10.3%)			0.46 (0.26, 0.84)	0.011	0.017	
CA+AA					0.65 (0.48, 0.88)	0.006	0.017	
rs2282679			7.61	0.022				0.584
AA	126 (54.6%)	510 (44.7%)			Reference			
CA	87 (37.7%)	513 (45.0%)			0.70 (0.51, 0.96)	0.029	0.029	
CC	18 (7.80%)	118 (10.3%)			0.51 (0.29, 0.91)	0.023	0.029	
CA+CC	. /	. ,			0.66 (0.49, 0.90)	0.008	0.024	

Table 2. Effect of GC gene SNPs on T2D risk

SNP: single-nucleotide polymorphism; OR: odds ratio; CI: confidence interval.

Data are expressed as the frequency and percentage (%).

 $^{\dagger}p$  values for chi-square test.

<sup>‡</sup>Adjusted for age, gender, smoking, drinking, exercise, married status, occupation, culture, family history of diabetes, and 25(OH)D3.

p values for odds ratio.

(Table 4). The rs4588 genotype was significantly associated with serum  $25(OH)D_3$  concentrations in the non-T2D group, and the AA genotype had a significantly higher serum  $25(OH)D_3$  concentrations than did those with the CC genotype (p=0.022). However, no significant association was found either between rs7041 and 25(OH)D<sub>3</sub> or between rs2282679 and 25(OH)D<sub>3</sub> in any of the groups. In addition, the rs2282679 genotype was associated with serum DBP concentrations in the T2D group, showing that the CA genotype was associated with lower DBP concentrations compared with the AA genotype (p=0.020). And rs7041 and rs4588 variants had no significant association with DBP in either the T2D or non-T2D groups.

The association of serum vitamin D metabolite concentrations and T2D risk was also investigated (Supplementary Table 1). For both serum 25(OH)D<sub>3</sub> and DBP, the risk of T2D in quartile 2 (Q2) was significantly higher than those in Q4 (OR=1.92, 95% CI: 1.20-3.06; OR=1.76, 95% CI: 1.12-2.76, respectively), after adjusting for age, gender, married status, education, smoking status, drinking status, physical activity, family history of diabetes.

#### DISCUSSION

Vitamin D is related to the pathogenesis of T2D, and the GC gene, as the most important transporter of vitamin D, plays a regulatory role in vitamin D metabolism. Exploring the relationship between GC SNPs and T2D as well as vitamin D may provide valuable information for the prediction, occurrence, and development of T2D. Our study indicates that GC variants (rs7041, rs4588, and rs2282679) are associated with T2D risk in a Chinese rural population. It was also found that lower vitamin D concentrations were associated with a higher T2D risk. Furthermore, we found that rs4588 and rs2282679 genotypes were associated with vitamin D metabolite concentrations.

Several studies have investigated the association of GC gene polymorphisms and the risk of T2D with conflicting results.<sup>13,15,16,20</sup> Zhao et al demonstrated a significant dif-

ference between rs7041 polymorphism and T2D incidence.<sup>21</sup> Ye et al and Malecki et al did not find evidence for an association between GC gene SNPs and T2D in French Caucasians and Poland.<sup>15,16</sup> Rahman et al and Hirai et al identified that there was an association of GC SNPs and T2D in Bangladesh and Japan respectively.<sup>13,20</sup> In our study, we found that rs7041, rs4588, and rs2282679 polymorphisms within the GC gene were all significantly associated with risk of T2D in the Chinese rural population. Moreover, our results revealed that the mutation of rs7041 was associated with a higher risk of T2D, while the mutations of rs2282679 and rs4588 were a protective factor for the progression of T2D. It has been reported that beta cells behave abnormally when islet fatty acid concentration increases. Therefore, we speculate that GC gene variants may alter fatty acid concentrations and thus affect the development of diabetes.<sup>22</sup> In addition, results of subgroup analysis revealed that the mutation of rs4588 and rs2282679 were associated with a lower risk of T2D in females, subjects younger than 60 years and without a family history of diabetes. To the best of our knowledge, gender, genetic factors, and age have been considered high-risk factors for T2D. Male has a higher risk of T2D than female at lower age and body mass index.<sup>23</sup> Relevant reports indicated a parental history of diabetes was related to a 2-6 times increase in diabetes risk.<sup>24,25</sup> Compared with younger ages, diabetes was more prevalent among older ages (60-79 years).26,27 No significant difference was found among male, subjects over 60 years and with a family history of diabetes, which can be explained by the distinct effects of gender, age, and family history of diabetes on T2D may blur the association between GC SNPs and T2D risk. In addition, we found that rs7041 was associated with an increased T2D risk in subjects without hypertension. In our study, most patients with hypertension have taken antihypertensive drugs such as calcium channel blockers, angiotensin-converting enzyme inhibitors, or angiotensin-receptor blockers, which may decrease the incidence of diabetes.28 This could interfere with genetic effects on the progression of T2D

Variables	T2D	Non-T2D -	rs7041		rs4588		rs2282679	
			OR (95% CI) <sup>†</sup>	$p^{\ddagger}$	OR (95% CI) <sup>†</sup>	$p^{\ddagger}$	OR (95% CI) <sup>†</sup>	$p^{\ddagger}$
Age (years)					· · · · · ·			
<60	107 (7.80%)	780 (56.9%)	1.50 (0.96, 2.32)	0.073	0.57 (0.37, 0.89)	0.013	0.57 (0.37, 0.88)	0.012
≥60	124 (9.00%)	361 (26.3%)	1.13 (0.73, 1.75)	0.597	0.75 (0.48, 1.17)	0.205	0.78 (0.50, 1.22)	0.274
Gender	× /	. ,						
Male	96 (7.00%)	580 (42.3%)	1.38 (0.87, 2.18)	0.173	0.69 (0.44, 1.10)	0.121	0.72 (0.45, 1.14)	0.162
Female	134 (9.80%)	561 (40.9%)	1.13 (0.75, 1.69)	0.558	0.63 (0.42, 0.94)	0.029	0.62 (0.41, 0.93)	0.022
Family history of diabetes	. ,	. ,						
Yes	53 (3.90%)	264 (19.2%)	2.07 (1.03, 4.16)	0.040	0.80 (0.40, 1.57)	0.512	0.87 (0.44, 1.70)	0.674
No	174 (12.7%)	869 (63.3%)	1.08 (0.76, 1.52)	0.667	0.65 (0.46, 0.92)	0.016	0.65 (0.46, 0.91)	0.013
Hypertension	. ,	. ,						
Yes	133 (9.70%)	450 (32.8%)	1.00 (0.67, 1.50)	0.993	0.63 (0.42, 0.97)	0.032	0.68 (0.46, 1.03)	0.053
No	94 (6.90%)	685 (49.9%)	1.62 (1.02, 2.56)	0.041	0.50 (0.31, 0.80)	0.004	0.46 (0.28, 0.74)	0.001
VDD+VDI								
Yes	171 (12.5%)	700 (51.0%)	1.40 (0.98, 1.98)	0.062	0.67 (0.47, 0.96)	0.027	0.69 (0.49, 0.99)	0.045
No	60 (4.40%)	441 (32.1%)	0.99 (0.55, 1.82)	0.995	0.54 (0.30, 0.97)	0.039	0.52 (0.28, 0.95)	0.035

Table 3. Effect of GC gene SNPs on T2D risk stratified by subgroups

SNP: single-nucleotide polymorphism; VDD: Vitamin D deficiency; VDI: vitamin D insufficient; OR: odds ratio; CI: confidence interval.

Data are expressed as the frequency and percentage (%).

<sup>†</sup>Adjusted for age, gender, smoking, drinking, exercise, married status, occupation, culture, family history of diabetes, and 25(OH)D<sub>3</sub> under the dominant inheritance model. <sup>\*</sup>*p* values for odds ratio.

SNPs		25(OH)D <sub>3</sub> (ng/mL) <sup>†</sup>		$\chi^2$	p value <sup>‡</sup>		DBP (ng/mL) <sup>†</sup>		$\chi^2$	p value <sup>‡</sup>
rs7041	TT	TG	GG			TT	TG	GG		
T2D	18.5 (15.5, 37.5)	17.2 (14.4, 28.5)	20.1 (16.3, 37.1)	4.07	0.131	125 (99.2, 276)	117 (97.7, 183)	147 (105, 199)	2.29	0.318
non-T2D	21.1 (15.2, 52.3)	20.3 (14.9, 45.3)	26.0 (16.9, 50.5)	2.63	0.269	154 (102, 412)	153 (102, 353)	171 (110, 423)	1.52	0.467
rs4588	CC	CA	AA			CC	CA	AA		
T2D	19.1 (15.0, 38.3)	17.5 (14.6, 29.6)	17.5 (16.8, 23.1)	1.61	0.446	137 (102, 285)	112 (96.2, 198)	122 (100, 252)	5.40	0.067
non-T2D	23.7 (15.3, 51.5)	20.4 (14.7, 44.8)	26.8 (15.9, 58.8)	7.77	0.022	160 (103, 424)	143 (101, 361)	180 (111, 416)	4.46	0.097
rs2282679	AA	CA	CC			AA	CA	CC		
T2D	19.6 (15.2, 39.5)	17.0 (14.4, 28.2)	19.8 (16.8, 37.5)	4.86	0.088	137 (103, 323)	112 (96.1, 183)	123 (100, 252)	7.85	0.020
non-T2D	22.2 (15.4, 52.3)	20.4 (14.6, 44.8)	21.6 (15.8, 54.1)	3.76	0.153	162 (104, 432)	144 (101, 367)	166 (111, 379)	3.77	0.151

Table 4. Distribution of vitamin D metabolites under different genotypes

SNP: single-nucleotide polymorphism; DBP: Vitamin D binding protein.

<sup>†</sup>Data are expressed as medians (interquartile ranges) for continuous variables with non-normal distributions.

\*Kruskal-Wallis one-way analysis was performed to evaluate the association between genotypes and vitamin D metabolites concentrations.

among these patients.

Increasing evidence has suggested that vitamin D has a protective effect on T2D and high 25(OH)D<sub>3</sub> concentrations in the body are associated with a lower risk of T2D.<sup>29-31</sup> As the main protein carrier of serum  $25(OH)D_3$ , DBP is considered to be the main dominant factor of 25(OH)D<sub>3</sub> concentrations. Therefore, simultaneous measurement of DBP concentrations may reveal a causal pathway between 25(OH)D<sub>3</sub> and the risk of T2D. Our study discovered that the concentrations of serum 25(OH)D<sub>3</sub> and DBP in the T2D group were significantly lower than in the non-T2D group, which is consistent with the results of other studies.<sup>32-35</sup> Additionally, we found that both lower concentrations of 25(OH)D<sub>3</sub> and DBP were associated with an increased risk of T2D compared with the highest quartile. The relationship between vitamin D and T2D may be explained by the following mechanisms. First, vitamin D can regulate insulin secretion by maintaining calcium pool balance in vitro and in vivo.36 Second, vitamin D can reduce the production and release of inflammatory factors, inhibit the low-grade inflammatory response, and increase the expression of insulin receptor genes in insulin resistance target organs (fat, skeletal muscle, and liver), thereby improving the tissue response to insulin.37 Third, vitamin D may suppress the secretion of parathyroid hormone in the body and the renal angiotensin aldosterone system, thereby improving insulin sensitivity.<sup>38-40</sup> In conclusion, vitamin D may affect the progression of T2D by regulating insulin secretion and insulin resistance.

Sedky et al. and Zhang et al also explored the association between the GC SNPs (rs7041, rs4588, and rs2282679) and serum  $25(OH)D_3$  concentrations, but they did not find a significant difference.<sup>11,41</sup> Contrarily, Ahn et al and Wang et al in their studies found that rs2282679 variants were associated with lower serum  $25(OH)D_3$ concentrations in white Europeans.<sup>8,9</sup> In our study, rs4588 genotypes were significantly associated with serum  $25(OH)D_3$  concentrations in the non-T2D group and rs2282679 genotype was associated with serum DBP



Figure 2. Graphical abstract.

concentrations in the T2D group. The results of our study may suggest a role of GC polymorphisms in regulating vitamin D metabolite concentrations and T2D risk. Previous studies have shown that 1,25 dihydroxy vitamin D<sub>3</sub> [1,25(OH)<sub>2</sub>D<sub>3</sub>], a metabolite of 25(OH)D<sub>3</sub>, influences insulin secretion and insulin resistance.42,43 The 1,25(OH)2D<sub>3</sub> could directly bind to vitamin D receptors (VDR) in pancreatic  $\beta$  cells, and stimulate the production of insulin by regulating gene transcription.<sup>44</sup> On the other hand, 1,25(OH)<sub>2</sub>D<sub>3</sub> could mediate the transcriptional activation of insulin receptor genes, increasing the number of insulin receptors on the surface of target cells, thus promoting insulin signaling and maintaining insulin sensitivity.45 Therefore, we hypothesized that GC variants could affect serum DBP concentrations, leading to changes in the concentrations of 1,25(OH)<sub>2</sub>D<sub>3</sub>, thus playing a critical role in the occurrence and development of T2D.

In summary, our study suggests that the risk of T2D may be influenced by the GC gene polymorphisms and serum  $25(OH)D_3$  concentrations. Vitamin D plays a pivotal role in insulin secretion and insulin sensitivity, and variations in the GC gene can affect the concentrations of vitamin D metabolites, which in turn affect the secretion of insulin, leading to disorders of glucose metabolism and even the occurrence of T2D (Figure 2). Our findings encourage further research to elucidate the mechanisms underlying T2D.

The advantage of the present study is that we found a significant association between the rs2282679 variant and T2D risk in a Chinese rural population. But limitations should be noted. First, this study was conducted in July and August each year when serum  $25(OH)D_3$  measured in summer may be higher due to sunlight and skin exposure. Second, given that our study included only the Chinese rural population, the conclusions observed in our research may not be directly extrapolated to other races. Third, we only focused on the GC gene involved in vitamin D metabolism. It is not clear whether there are other genes or factors that interact with the GC gene to cause T2D.

#### Conclusion

GC gene polymorphisms (rs7041, rs4588, and rs2282679) are associated with T2D risk in a Chinese rural population. Lower concentrations of 25(OH)D<sub>3</sub> and DBP were associated with a higher risk of T2D. Further, we found rs7041 and rs4588 were associated with 25(OH)D<sub>3</sub> and DBP concentrations. These findings highlight the importance of genetic polymorphisms in T2D progression and suggest that a complex combination of vitamin D metabolites and GC SNPs may underlie T2D susceptibility. Genetic studies could screen for risk genotypes to complement traditional diabetes risk factors, so identifying high-risk patients earlier.

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#### AUTHOR DISCLOSURES

The authors have no relevant interests to declare.

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#### REFERENCES

- Chen L, Magliano DJ, Zimmet PZ. The worldwide epidemiology of type 2 diabetes mellitus--present and future perspectives. Nat Rev Endocrinol. 2011;8:228-36. doi: 10. 1038/nrendo.2011.183.
- Saeedi P, Petersohn I, Salpea P, Malanda B, Karuranga S, Unwin N et al. Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: Results from the International Diabetes Federation Diabetes Atlas, 9(th) edition. Diabetes Res Clin Pract. 2019;157: 107843. doi: 10.1016/j.diabres.2019.107843.
- Wang Y, Duan L, Yu S, Liu X, Han H, Wang J, Li W. Association between "solute carrier family 30 member 8" (SLC30A8) gene polymorphism and susceptibility to type 2 diabetes mellitus in Chinese Han and minority populations: an updated meta-analysis. Asia Pac J Clin Nutr. 2018;27: 1374-90. doi: 10.6133/apjcn.201811\_27(6).0025.
- Li SX, Imamura F, Schulze MB, Zheng J, Ye Z, Agudo A et al. Interplay between genetic predisposition, macronutrient intake and type 2 diabetes incidence: analysis within EPIC-InterAct across eight European countries. Diabetologia. 2018;61:1325-32. doi: 10.1007/s00125-018-4586-2.
- Fawzy MS, Beladi FIA. Association of circulating vitamin D, VDBP, and vitamin D receptor expression with severity of diabetic nephropathy in a group of Saudi type 2 diabetes mellitus patients. Clin Lab. 2018;64:1623-33. doi: 10. 7754/Clin.Lab.2018. 180401.
- Karuwanarint P, Phonrat B, Tungtrongchitr A, Suriyaprom K, Chuengsamarn S, Schweigert FJ, Tungtrongchitr R. Vitamin D-binding protein and its polymorphisms as a predictor for metabolic syndrome. Biomark Med. 2018;12: 465-73. doi: 10.2217/bmm-2018-0029.
- Braun A, Bichlmaier R, Cleve H. Molecular analysis of the gene for the human vitamin-D-binding protein (groupspecific component): allelic differences of the common genetic GC types. Hum Genet. 1992;89:401-6. doi: 10. 1007/bf00194311.
- Ahn J, Yu K, Stolzenberg-Solomon R, Simon KC, McCullough ML, Gallicchio L et al. Genome-wide association study of circulating vitamin D levels. Hum Mol Genet. 2010;19:2739-45. doi: 10.1093/hmg/ddq155.
- Rivera-Paredez B, Hidalgo-Bravo A, de la Cruz-Montoya A, Martínez-Aguilar MM, Ramírez-Salazar EG, Flores M et al. Association between vitamin D deficiency and common variants of Vitamin D binding protein gene among Mexican Mestizo and indigenous postmenopausal women. J Endocrinol Invest. 2020;43:935-46. doi: 10.1007/s40618-019-01177-5.
- Jorde R, Schirmer H, Wilsgaard T, Bøgeberg Mathiesen E, Njølstad I, Løchen ML, Joakimsen RM, Grimnes G. The DBP phenotype Gc-1f/Gc-1f is associated with reduced risk of cancer. The Tromsø Study. PLoS One. 2015;10:e0126359. doi: 10.1371/journal.pone.0126359.
- Sedky NK, Hassanein SI, Gad MZ. Independent assortment of GC gene polymorphism (rs2282679) and 25hydroxyvitamin D levels in coronary artery disease. Can J Physiol Pharmacol. 2018;96:345-51. doi: 10.1139/cjpp-2017-0334.
- 12. Gezen-Ak D, Alaylıoğlu M, Genç G, Gündüz A, Candaş E, Bilgiç B et al. GC and VDR SNPs and vitamin D levels in Parkinson's disease: The relevance to clinical features. Neuromolecular Med. 2017;19:24-40. doi: 10.1007/s12017-016-8415-9.

- Rahman MM, Hosen MB, Faruk MO, Hasan MM, Kabir Y, Howlader MZH. Association of vitamin D and vitamin D binding protein (DBP) gene polymorphism with susceptibility of type 2 diabetes mellitus in Bangladesh. Gene. 2017;636:42-7. doi: 10.1016/j.gene.2017.09.008.
- 14. Wang G, Li Y, Li L, Yu F, Cui L, Ba Y, Li W, Wang C. Association of the vitamin D binding protein polymorphisms with the risk of type 2 diabetes mellitus: a meta-analysis. BMJ Open. 2014;4:e005617. doi: 10.1136/ bmjopen-2014-005617.
- 15. Ye WZ, Dubois-Laforgue D, Bellanné-Chantelot C, Timsit J, Velho G. Variations in the vitamin D-binding protein (Gc locus) and risk of type 2 diabetes mellitus in French Caucasians. Metabolism. 2001;50:366-9. doi: 10.1053/meta. 2001.20172.
- 16. Malecki MT, Klupa T, Wanic K, Cyganek K, Frey J, Sieradzki J. Vitamin D binding protein gene and genetic susceptibility to type 2 diabetes mellitus in a Polish population. Diabetes Res Clin Pract. 2002;57:99-104. doi: 10.1016/s0168-8227(02)00020-7.
- Alberti KG, Zimmet PZ. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. Diabet Med. 1998;15:539-53. doi: 10.1002/(sici)1096-9136(199807)15: 7<539::Aid-dia668>3.0.Co;2-s.
- Kotlarczyk MP, Perera S, Ferchak MA, Nace DA, Resnick NM, Greenspan SL. Vitamin D deficiency is associated with functional decline and falls in frail elderly women despite supplementation. Osteoporos Int. 2017;28:1347-53. doi: 10. 1007/s00198-016-3877-z.
- Fawzy MS, Toraih EA, Al Ageeli E, Mohamed AM, Abu AlSel BT, Kattan SW, Alelwani W. Group-specific component exon 11 haplotypes (D432E and T436K) and risk of albuminuria in type 2 diabetes mellitus patients. Arch Physiol Biochem. 2022;128:111-20. doi: 10.1080/13813455. 2019.1665689.
- Hirai M, Suzuki S, Hinokio Y, Chiba M, Kasuga S, Hirai A, Toyota T. Group specific component protein genotype is associated with NIDDM in Japan. Diabetologia. 1998;41: 742-3. doi: 10.1007/s001250050980.
- 21. Zhao Y, Wang G, Li Y, Liu X, Liu L, Yang K, Wang C, Wei S. Evaluation of the associations of GC and CYP2R1 genes and gene-obesity interactions with type 2 diabetes risk in a Chinese rural population. Ann Nutr Metab. 2020;76: 175-82. doi: 10.1159/000508024.
- Unger RH. Lipotoxicity in the pathogenesis of obesitydependent NIDDM. Genetic and clinical implications. Diabetes. 1995;44:863-70. doi: 10.2337/diab.44.8.863.
- 23. Kautzky-Willer A, Harreiter J, Pacini G. Sex and gender differences in risk, pathophysiology and complications of type 2 diabetes mellitus. Endocr Rev. 2016;37:278-316. doi: 10.1210/er.2015-1137.
- Harrison TA, Hindorff LA, Kim H, Wines RC, Bowen DJ, McGrath BB, Edwards KL. Family history of diabetes as a potential public health tool. Am J Prev Med. 2003;24:152-9. doi: 10.1016/ s0749-3797(02)00588-3.
- Tsenkova VK, Karlamangla AS, Ryff CD. Parental history of diabetes, positive affect, and diabetes risk in adults: Findings from MIDUS. Ann Behav Med. 2016;50:836-43. doi: 10.1007/s12160-016-9810-z.
- Chang AM, Halter JB. Aging and insulin secretion. Am J Physiol Endocrinol Metab. 2003;284:E7-12. doi: 10.1152/ ajpendo.00366.2002.
- 27. Hosseini Z, Whiting SJ, Vatanparast H. Type 2 diabetes prevalence among Canadian adults dietary habits and

sociodemographic risk factors. Appl Physiol Nutr Metab. 2019;44:1099-104. doi: 10.1139/apnm-2018-0567.

- Aksnes TA, Reims HM, Kjeldsen SE, Mancia G. Antihypertensive treatment and new-onset diabetes mellitus. Curr Hypertens Rep. 2005;7:298-303. doi: 10.1007/s11906-005-0029-9.
- Martínez-Sena T, Soluyanova P, Guzmán C, Valdivielso JM, Castell JV, Jover R. The vitamin D receptor regulates glycerolipid and phospholipid metabolism in human hepatocytes. Biomolecules. 2020;10:493. doi: 10.3390/ biom10030493.
- Ekmekcioglu C, Haluza D, Kundi M. 25-hydroxyvitamin D status and risk for colorectal cancer and type 2 diabetes mellitus: A systematic review and meta-analysis of epidemiological studies. Int J Environ Res Public Health. 2017;14:127. doi: 10.3390/ijerph14020127.
- 31. Lucato P, Solmi M, Maggi S, Bertocco A, Bano G, Trevisan C et al. Low vitamin D levels increase the risk of type 2 diabetes in older adults: A systematic review and meta-analysis. Maturitas. 2017;100:8-15. doi: 10.1016/j.maturitas. 2017.02.016.
- 32. Shen L, Zhuang QS, Ji HF. Assessment of vitamin D levels in type 1 and type 2 diabetes patients: Results from metaanalysis. Mol Nutr Food Res. 2016;60:1059-67. doi: 10. 1002/mnfr.201500937.
- 33. Afzal S, Bojesen SE, Nordestgaard BG. Low 25hydroxyvitamin D and risk of type 2 diabetes: a prospective cohort study and metaanalysis. Clin Chem. 2013;59:381-91. doi: 10.1373/clinchem. 2012.193003.
- 34. Parveen R, Kapur P, Venkatesh S, Agarwal NB. Attenuated serum 25-hydroxyvitamin D and vitamin D binding protein associated with cognitive impairment in patients with type 2 diabetes. Diabetes Metab Syndr Obes. 2019;12:1763-72. doi: 10.2147/ dmso.S207728.
- 35. Jain SK, Kahlon G, Bass P, Levine SN, Warden C. Can Lcysteine and vitamin D rescue vitamin D and vitamin D binding protein levels in blood plasma of African American type 2 diabetic patients? Antioxid Redox Signal. 2015;23: 688-93. doi: 10.1089/ars.2015.6320.
- 36. Mitri J, Dawson-Hughes B, Hu FB, Pittas AG. Effects of vitamin D and calcium supplementation on pancreatic  $\beta$  cell function, insulin sensitivity, and glycemia in adults at high

risk of diabetes: the Calcium and Vitamin D for Diabetes Mellitus (CaDDM) randomized controlled trial. Am J Clin Nutr. 2011;94:486-94. doi: 10.3945/ajcn. 111.011684.

- Wimalawansa SJ. Associations of vitamin D with insulin resistance, obesity, type 2 diabetes, and metabolic syndrome. J Steroid Biochem Mol Biol. 2018;175:177-89. doi: 10. 1016/j.jsbmb.2016. 09.017.
- 38. Li YC, Kong J, Wei M, Chen ZF, Liu SQ, Cao LP. 1,25-Dihydroxyvitamin D(3) is a negative endocrine regulator of the renin-angiotensin system. J Clin Invest. 2002;110:229-38. doi: 10.1172/jci15219.
- Chiu KC, Chuang LM, Lee NP, Ryu JM, McGullam JL, Tsai GP, Saad MF. Insulin sensitivity is inversely correlated with plasma intact parathyroid hormone level. Metabolism. 2000;49:1501-5. doi: 10.1053/meta.2000.17708.
- 40. Engeli S, Schling P, Gorzelniak K, Boschmann M, Janke J, Ailhaud G, Teboul M, Massiéra F, Sharma AM. The adipose-tissue renin-angiotensin-aldosterone system: role in the metabolic syndrome? Int J Biochem Cell Biol. 2003;35: 807-25. doi: 10.1016/s1357-2725(02)00311-4.
- 41. Zhang Y, Yang S, Liu Y, Ren L. Relationship between polymorphisms in vitamin D metabolism-related genes and the risk of rickets in Han Chinese children. BMC Med Genet. 2013;14:101. doi: 10.1186/1471-2350-14-101.
- 42. Yu F, Cui LL, Li X, Wang CJ, Ba Y, Wang L, Li J, Li C, Dai LP, Li WJ. The genetic polymorphisms in vitamin D receptor and the risk of type 2 diabetes mellitus: an updated meta-analysis. Asia Pac J Clin Nutr. 2016;25:614-24. doi: 10.6133/apjcn.092015.12.
- 43. Kadowaki S, Norman AW. Demonstration that the vitamin D metabolite 1,25(OH)2-vitamin D3 and not 24R,25(OH)2vitamin D3 is essential for normal insulin secretion in the perfused rat pancreas. Diabetes. 1985;34:315-20. doi: 10. 2337/diab.34.4.315.
- 44. Chuang JC, Cha JY, Garmey JC, Mirmira RG, Repa JJ. Research resource: nuclear hormone receptor expression in the endocrine pancreas. Mol Endocrinol. 2008;22:2353-63. doi: 10.1210/me.2007-0568.
- 45. Szymczak-Pajor I, Drzewoski J, Śliwińska A. The molecular mechanisms by which vitamin D prevents insulin resistance and associated disorders. Int J Mol Sci. 2020;21:6644. doi: 10.3390/ ijms21186644.

	The concentration of vitamin D metabolites						
	Q1†‡	Q2 <sup>†‡</sup>	Q3†‡	Q4†‡			
25(OH)D <sub>3</sub> (ng/ml)							
model 1 <sup>§</sup>	1.59 (0.99, 2.56)	1.92 (1.21, 3.06)	1.64 (1.02, 2.63)	Reference			
model 2¶	1.59 (0.97, 2.60)	2.14 (1.32, 3.46)	1.69 (1.04, 2.77)	Reference			
DBP (ng/ml)							
model 1§	1.71 (1.07, 2.72)	1.85 (1.16, 2.93)	1.20 (0.74, 1.96)	Reference			
model 2¶	1.76 (1.09, 2.83)	1.82 (1.13, 2.93)	1.20 (0.73, 1.98)	Reference			

Supplementary table 1. Odds ratios and 95% confidence intervals for T2D by quartile of vitamin D metabolites

DBP: Vitamin D binding protein.

<sup>†</sup>25(OH)D<sub>3</sub> (ng/ml) – Q1 (<15.3); Q2 (15.3–21.0); Q3 (21.0–45.6); Q4 ( $\geq$ 45.6). <sup>‡</sup>DBP (ng/ml) – Q1 (<104); Q2 (104–149); Q3 (149–368); Q4 ( $\geq$ 368).

<sup>§</sup>Model 1 adjusted for age, gender. <sup>¶</sup>Model 2 adjusted for age, gender, smoking, drinking, exercise, married status, occupation, culture, and family history of diabetes.



Supplementary figure 1. The results of linkage disequilibrium between the three SNPs.