Original Article

Associations of postprandial lipemia with trunk/leg fat ratio in young normal weight women independently of fat mass and insulin resistance

Mika Takeuchi NRD, PhD¹, Ayaka Tsuboi NRD^{2,3}, Miki Kurata NRD^{1,2}, Tsutomu Kazumi MD, PhD^{2,4}, Keisuke Fukuo MD, PhD^{1,2}

¹Department of Food Sciences and Nutrition, School of Human Environmental Sciences, Nishinomiya, Japan ²Research Institute for Nutrition Sciences, Mukogawa Women's University, Nishinomiya, Hyogo, Japan ³Department of Nutrition, Osaka City Juso Hospital, Osaka, Japan ⁴Diabetes Division, Kohnann Kakogawa Hospital, Kakogawa, Hyogo, Japan

Background and Objectives: To determine whether postprandial lipemia is associated with fat distribution even in young, normal weight women independently of fat mass, adipokines, insulin resistance and systemic inflammation. **Methods and Study Design:** Female college students (ages 21-24, n=35) underwent dual-energy X-ray absorptiometry and a standardized breakfast providing 17 g triglycerides (TG). Serum lipids, lipoproteins, apolipoproteins, adipokines and markers of insulin resistance and inflammation were measured in fasting blood samples. **Results:** In crude analyses, postprandial lipemia, as assessed by 0-2 h area under the curve of serum TG (TG-AUC), was positively associated with fasting TG, trunk/leg fat ratio, apolipoprotein B, leptin/adiponectin ratio and log high-sensitivity CRP. Multiple linear regression analysis with these 5 variables as independent variables revealed that fasting TG (p<0.001) and trunk/ leg fat ratio (p=0.001), were independent positive predictors of TG-AUC (R²=0.923). Women with high compared to low TG-AUC were characterized by higher trunk/leg fat ratio, elevated apolipoprotein B and leptin/adiponectin ratio. **Conclusion:** Trunk/leg fat ratio, a marker of central adiposity, is a significant predictor of postprandial lipemia even in young women who are normal weight and insulin-sensitive, suggesting a modifiable pathway to postprandial hypertriglyceridemia, a cardiometabolic risk factor. These findings should be confirmed in studies employing more participants.

Key Words: postprandial triglyceridemia, trunk fat, leg fat, trunk/leg fat ratio, insulin resistance

INTRODUCTION

High levels of serum triglycerides (TG) play a role in cardiovascular disease.¹ New insights strongly suggest that TG-rich lipoproteins represent causal risk factors for low-grade inflammation, atherosclerotic cardiovascular disease, and all-cause mortality.² TG levels are commonly measured in the fasting state; TG levels, however, increase significantly postprandially, and an important role in the pathogenesis of atherosclerosis-related diseases has been postulated for postprandial lipids. Post-alimentary lipemia was first described as an atherogenic phenomenon by Zilversmit.³ Later, it received more attention after the discovery that TG-rich lipoproteins are atherogenic.⁴ Postprandial abnormalities in TG metabolism (increased plasma TG concentrations and prolonged postprandial response after a fatty meal) are a hallmark of patients with established cardiovascular disease.⁵ As recently reviewed,^{6,7} the postprandial lipemic response is influenced by numerous factors including background dietary pattern and meal composition, lifestyle conditions (smoking, alcohol consumption and physical activity), physiological factors (age, gender and menopause), pathological conditions (obesity, metabolic syndrome and type 2 diabetes) and genetic factors.

It has been reported that obesity in general,⁸ abdominal obesity in particular,⁹⁻¹² is associated with postprandial lipemia. Many investigators have shown repeatedly that fasting TG is the major determinant of postprandial lipemia.^{9,10,13} In the present study, therefore, we tested the hypothesis that the ratio of trunk fat to leg fat, measured using whole-body dual-energy X-ray absorptiometry (DXA), is a predictor of postprandial lipemia independent of fat mass, fasting TG, insulin resistance and inflammation even in young female college students, a population in which confounding factors are so scarce.¹⁴

PARTICIPANTS AND METHODS

Female collegiate athletes and untrained female students (n=17 and 18, respectively, aged 21-24 years) were re-

Corresponding Author: Dr Tsutomu Kazumi, Research Institute for Nutrition Sciences, Mukogawa Women's University, 6-46, Ikebiraki-cho, Nishinomiya, Hyogo, 663-8558, Japan. Tel: +81-798-45-9038; Fax: +81-798-45-3566 Email: kazumi@mukogawa-u.ac.jp Manuscript received 13 June 2016. Initial review completed 8 November 2016. Revision accepted 16 November 2016. doi: 10.6133/apjcn.062017.02 cruited as volunteers. Athletes and non-athletes were students from the Department of Health and Sports Sciences and the Department of Food Sciences and Nutrition of the Mukogawa Women's University, respectively. Athletes had been training regularly for 2 years or longer prior to the study, 5 h a day, and 6 days a week and participated regularly in competitive events in their respective sport specialties. Non-athletes were not engaged in any regular sport activity. Subjects with clinical diagnosed acute or chronic inflammatory diseases, endocrine, cardiovascular, hepatic, renal diseases, hormonal contraception, unusual dietary habits were excluded. Nobody reported to receive any medications or have regular supplements. The study was approved by the Ethics Committees of the University (No. 07-28) to be in accordance with the Helsinki declaration. All subjects gave written consent after the experimental procedure had been explained.

After 12 h overnight fasting, participants underwent a standardized meal test as described later and measurements of anthropometric indices and body composition in the morning, as previously described.¹⁴ Athletes had regular training on the day before measurements. In fasted blood samples, the following was measured as previously reported;¹⁴ plasma glucose, serum insulin, triglycerides, cholesterol, HDL cholesterol, apolipoprotein A-1 (apoA1), apolipoprotein B-100 (apoB), non-esterified fatty acid (NEFA), liver enzymes, adiponectin, leptin, high- sensitivity C-reactive protein (hsCRP), tumor necrosis factor- α (TNF- α), plasminogen activator inhibitor-1 (PAI-1). Insulin resistance was determined by homeostasis model assessment (HOMA-IR)¹⁵ and leptin/adiponectin ratio.¹⁶ Serum concentrations of hsCRP and TNF- α below the limit of detection were assigned a value of 20 µg/mL and 0.50 pg/ml (the lowest limit of detection), respectively.

Lean tissue mass, fat mass, and bone mineral mass for arms, legs, trunk and the total body were measured using whole-body DXA (Hologic QDR-2000, software version 7.20D, Waltham, MA) as previously reported.¹⁴ Although DXA cannot distinguish between intraabdominal and subcutaneous abdominal fat, research showed strong correlations between trunk fat mass measured with DXA and intraabdominal fat measured with CT.¹²

The standardized test meal was developed by the Japanese Diabetes Society to assess both postprandial hyperglycemia and hyperlipemia. This meal was composed as a breakfast meal (total energy 450 kcal) and provided 33.3% of calories from fat (16.7 g), 51.4% from carbohydrates (57.8 g), and 15.3% from protein (17.2 g). The test meal contained more fat than a typical Japanese breakfast (20-25%) but comparable energy content (median: 423 kcal). Participants were asked to consume the entire meal within 15 min. Venous blood was drawn at baseline (0 min), 30, 60, and 120 min after the start of the meal for the measurement of plasma glucose, serum insulin, TG and NEFA concentrations. The area under the concentration curve (AUC) was calculated with the trapezoidal method. Meal-stimulated insulin secretion (MSIS) was calculated as follows;

MSIS = (insulin at 30 min-fasting insulin)/ (glucose at 30 min-fasting glucose), where insulin in μ U/mL and glucose in mg/dL.

Data were presented as mean \pm SD unless otherwise

stated. Due to deviation from normal distribution, HOMA-IR and inflammatory markers were logarithmically transformed for analysis. Correlation coefficients of TG AUC with cardiometabolic parameters were evaluated by Pearson correlation analysis. Stepwise multiple linear regression analyses were performed to further identify the most significant variables contributing to the variation of TG-AUC. A two-tailed p<0.05 was considered statistically significant. All calculations were performed with SPSS system 15.0 (SPSS Inc, Chicago, IL).

RESULTS

As previously reported,^{14,17} participants in total were normal weight, normolipidemic and normoglycemic (Table 1). They were also insulin-sensitive as demonstrated by HOMA-IR of 0.76. Athletes compared to non-athletes had lower percentage fat in any region measured in the present study. Although fat mass in arms and legs did not differ between the 2 groups, trunk fat and trunk/leg fat ratio were tended to be higher in athletes. The 2 groups did not differ in TG at 4 time points during a meal test, TG-AUC, HDL cholesterol and apolipoprotein A1 and B. Athletes compared to non-athletes had lower leptin but comparable adiponectin and hence lower leptin/adiponectin ratio. Although glucose-AUC did not differ between groups, insulin-AUC was lower in athletes, suggesting lower muscle insulin sensitivity in athletes.

When analyzed as a whole or separately (Table 2), TG-AUC showed a positive association with a ratio of trunk fat to leg fat whereas no association was found with BMI, waist circumference, fat mass and percentage fat of any region studied. TG responses were strongly and positively associated with fasting TG and TG at 30, 60 and 120 min in meal tests (data not shown) and apoB whereas there was no association with HDL cholesterol and apoA1. As a whole group, TG-AUC was positively associated with leptin/adiponectin ratio and log hsCRP but not with glucose, insulin, HOMA-IR, serum adiponectin, leptin, TNF- α and PAI-1.

We conducted multiple linear regression analysis for TG AUC as a dependent variable (Table 3, model A). The model included 5 variables as independent variables, which showed significant associations with postprandial lipemia in univariate analysis. Fasting TG emerged as the strongest determinant and explained 88.9% of variations of postprandial lipemia. In addition to fasting TG, trunk/ leg fat ratio emerged as an independent determinant of postprandial triglyceridemia and the 2 variables explained 92.3% of variabilities of postprandial lipemia. Including total body fat, leg fat and trunk fat into the model as independent variables did not change the results (data not shown).

In order to confirm the association between trunk/leg fat ratio and TG-AUC, participants were stratified into 2 groups according to the median TG-AUC (133 mg/dL. 2h) (Table 4). Young women with high compared to low TG-AUC had higher trunk fat but comparable leg fat, and consequently, higher trunk/leg fat ratio. They had elevated leptin/adiponectin ratio as the result of elevated leptin and comparable adiponectin. ApoB and TG concentrations at 4 time points were all substantially higher in women with high compared to low TG-AUC while there

	Total	Non-athlete	Athlete	n volue
	n=35	n=18	n=17	<i>p</i> -value
BMI (kg/m ²)	21.8±2.4	22.0±2.8	21.7±1.9	0.704
Waist circumference (cm)	73.8±6.3	73.5±6.9	74.1±5.7	0.766
Arm fat (kg)	1.2±0.6	1.4±0.7	1.1±0.6	0.121
Leg fat (kg)	6.1±2.1	6.5±2.5	5.6±1.5	0.211
Trunk fat (kg)	7.3±2.7	8.2±2.8	6.5±2.4	0.059
Body fat (kg)	15.2±5.2	16.7±5.7	13.6±4.2	0.083
Percentage arm fat (%)	22.5±8.7	26.4±8.3	18.4±7.3	0.005
Percentage leg fat (%)	29.1±6.7	32.5±6.9	25.5±4.3	0.001
Percentage trunk fat (%)	27.0±7.4	31.0±6.3	22.7±6.1	0.000
Percentage body fat (%)	26.3±6.6	29.9±6.1	22.5±5.0	0.000
Trunk/leg fat ratio	1.22±0.25	1.29±0.27	1.14±0.22	0.074
Cholesterol (mg/dL)	178±23	175±22	182 ± 24	0.373
Fasting TG (mg/dL)	55±19	54±19	55±19	0.827
TG 30 min (mg/dL)	61±23	62±25	60±21	0.874
TG 60 min (mg/dL)	73±26	76±29	70±23	0.465
TG 120 min (mg/dL)	79±30	78±26	81±34	0.830
TG-AUC (mg/dL. 2 hr)	139±47	141±49	137±46	0.805
HDL cholesterol (mg/dL)	74±11	73±14	75±8	0.511
Apolipoprotein A1 (mg/dL)	159±14	155±16	164±12	0.080
Apolipoprotein B (mg/dL)	67±14	64±13	69±14	0.359
PAI-1 (ng/mL)	29±20	33±27	26±9	0.303
TNF- α (pg/mL)	0.63 ± 0.20	0.65 ± 0.26	0.60±0.11	0.409
Leptin (ng/mL)	7.0±4.3	9.0±4.7	4.9±2.6	0.003
Adiponectin (µg/mL)	11.2±4.5	11.3±4.3	11.1±4.8	0.923
Leptin/adiponectin ratio	0.72 ± 0.55	0.92 ± 0.63	0.51±0.36	0.025
IL-6 (pg/mL)	0.7±0.3	0.7±0.3	0.7±0.3	0.869
$hsCRP(\mu g/dL)$	42±91	31±50	54±121	0.471
Fasting glucose (mg/dL)	84±4	82±4	85±4	0.033
Fasting insulin (µU/mL)	3.7±2.0	4.1±2.0	3.3±2.1	0.258
HbA1c (%)	5.2±0.2	5.1±0.2	5.2±0.2	0.202
HOMA-IR	0.76±0.43	0.83±0.41	0.69±0.44	0.349
Glucose-AUC (mg/dL. 2 hr)	174±20	174 ± 20	175±21	0.852
Insulin-AUC (µU/mL. 2 hr)	44±23	54±24	34±17	0.009

Table 1. Anthropometric and biochemical characteristics of your	g women and serum triglyceride responses to stand-
ardized meal	

HOMA: homeostasis model assessment; IR: insulin resistance; AUC: area under the curve; PAI-1; plasminogen activator inhibitor-1; TG: triglycerides; TNF-α; tumor necrosis factor-α; IL-6; interleukin-6; hsCRP; high-sensitivity C-reactive protein. Mean±SD

was no difference in HDL cholesterol and apoA1 between the 2 groups. Markers of inflammation did not differ between groups.

We have done another analysis on associations of fat distribution and triglyceridemia (Table 5). In simple correlation analyses, trunk/leg fat ratio showed strong and positive associations with not only fasting TG but also postprandial triglyceridemia and hence TG-AUC. In contrast, there was no association of triglyceridemia with leg fat and associations with trunk fat were weak. However, after mutual adjustment, leg fat was negatively and trunk fat was positively associated with not only fasting TG but also postprandial triglyceridemia and hence TG-AUC. .

Although there was no difference in glucose at baseline and 120 min, insulin concentrations at 4 time points and HOMA-IR, plasma glucose at 30 and 60 min and hence glucose-AUC were lower in women with high compared to low TG-AUC. MSIS was calculated and logarithmically transformed. MSIS was tended to be higher (3.6 ± 4.2 vs 1.3 ± 1.6 , p=0.052) and log MSIS was significantly higher in women with high compared with low TG-AUC (0.30 ± 0.50 vs -0.04 ± 0.31 , p=0.026). Association between TG-AUC and log MSIS was significant in the total group (r=0.36, p<0.05) and in women with high TG-AUC (r=0.69, p<0.01) but not in those with low TG-AUC (r=0.08, NS). When log MSIS was added as an independent variable to model A of multiple linear regression analysis, log MSIS emerged as a significant determinant of TG-AUC (Table 3, model B).

DISCUSSION

The present study has shown that even in healthy, normal weight women in early adult life postprandial lipemia was associated with fasting TG, trunk/leg fat ratio, apoB, leptin/adiponectin ratio, which would reflect compromised adipose tissue function and provide a useful index of insulin action,¹⁶ and hsCRP, a marker of systemic lowgrade inflammation. Among those, fasting TG and trunk/ leg fat ratio were independent determinants of postprandial triglyceridemia. Women with high compared to low TG-AUC were characterized by higher trunk/leg fat ratio, elevated apolipoprotein B and leptin/adiponectin ratio. It should be emphasized that this investigation was conducted in 35 young, normal weight normolipidemic women, a population in which confounding factors are so scarce.^{14,17}

Positive association between diurnal triglyceridemia and waist circumference, a crude marker of abdominal fat

Table 2. Correlation coefficients of the area under the curve of triglycerides (TG-AUC)

	Whole	Non-athletes	Athletes
BMI (kg/m ²)	0.26	0.37	0.07
Waist circumference (cm)	0.12	0.20	0.01
Arm fat (kg)	0.18	0.18	0.17
Leg fat (kg)	0.05	0.17	-0.20
Trunk fat (kg)	0.33	0.42	0.22
Body fat (kg)	0.22	0.30	0.08
% arm fat (%)	0.16	0.11	0.21
% leg fat (%)	0.03	0.00	0.01
% trunk fat (%)	0.28	0.34	0.29
% body fat (%)	0.18	0.19	0.20
Trunk/leg fat ratio	0.55**	0.54^{*}	0.60^{*}
Fasting TG (mg/dL)	0.92***	0.97^{***}	0.88^{***}
HDL cholesterol (mg/dL)	-0.20	-0.28	-0.06
apolipoprotein A1 (mg/dL)	0.01	-0.09	0.20
apolipoprotein B (mg/dL)	0.65***	0.72^{**}	0.62^{*}
PAI-1 (ng/mL)	0.11	0.09	0.18
TNF- α (pg/mL)	0.30	0.47^{*}	-0.11
Leptin (ng/mL)	0.29	0.22	0.52^{*}
Adiponectin (µg/mL)	-0.31	-0.41	-0.22
Leptin/adiponectin ratio	0.40^{*}	0.39	0.49^{*}
IL-6 (pg/mL)	-0.06	-0.17	0.12
Log hsCRP	0.35*	0.45	0.26
Fasting glucose (mg/dL)	0.07	0.09	0.08
Fasting insulin (µU/mL)	0.28	0.30	0.25
HbA1c (%)	0.06	0.14	0.00
HOMA-IR	0.28	0.29	0.25
G-AUC (mg/dL. 2 hr)	-0.31	-0.22	-0.40
I-AUC (μ U/mL. 2 hr)	0.13	-0.10	0.47

Abbreviations are the same as in Table 1.

p*<0.05; *p*<0.01; ****p*<0.001.

Table 3. Multiple linear regression analysis for area under the curve of triglycerides (TG-AUC) during meal tests as a dependent variable

	Standardized β	p values	Cumulative R ²	
Model A				
Fasting triglycerides	0.808	0.000	0.889	
Trunk/leg fat ratio	0.175	0.001	0.923	
Model B				
Fasting triglycerides	0.808	0.000	0.889	
Trunk/leg fat ratio	0.175	0.001	0.923	
log MSIS	0.097	0.039	0.930	

Abbreviations are the same as in Table 1.

Model A included variables which showed significant associations with TG-AUC as independent variables: fasting triglycerides, trunk/leg fat ratio/apoB, leptin/adiponectin ratio and log hsCRP. Model B included log meal-stimulated insulin secretion (MSIS) in addition to model A.

accumulation, has been demonstrated in lean and overweight middle-aged adults.¹⁰ Studies by computed tomography have shown positive association between postprandial lipemia and visceral adipose tissue accumulation in nonobese, normal glucose-tolerant middle-aged men,¹¹ in overweight/obese adults9 and in men with impaired glucose tolerance.¹² In the present study using DXA, a gold standard measure of body composition, postprandial triglyceridemia was associated not with trunk fat, but with the ratio of trunk to leg fat in crude analyses. These findings have a twofold significance. Even in young female college students who were normal weight and normolipidemic postprandial TG clearance was associated with trunk/leg fat ratio. Independent and opposite associations of trunk and leg fat depots with adipokines, inflammatory markers, and metabolic syndrome including fasting TG have been shown in studies employing DXA including ours.^{14,18,19} We confirmed and extended that young women showed the opposing associations between upper-body and lower-body adipose accumulation with not only fasting TG but also postprandial triglyceridemia and hence TG-AUC in the present study. Therefore, it is reasonable to assume that association of postprandial TG clearance with trunk/leg fat ratio in the present study could be combined effects of trunk and leg fat although the ratio of trunk to peripheral or leg fat ratio is a good marker of abdominal or central adiposity.²⁰

In the present study, women with higher TG-AUC had elevated apoB, higher trunk/leg fat ratio, a marker of abdominal fat accumulation,²⁰ and higher leptin/adiponectin ratio, a marker of insulin resistance.¹⁶ These findings may be in line with the observation that abnormalities in

	TG-A		
	Low	High	p values
	n=17	n=18	
BMI (kg/m^2)	20.9±1.6	22.7±2.8	0.029
Waist circumference (cm)	72.1±3.9	75.4±7.6	0.124
Arm fat (kg)	1.1±0.5	1.4±0.7	0.096
Leg fat (kg)	5.7±1.2	6.4±2.6	0.327
Trunk fat (kg)	6.2±1.5	8.4±3.2	0.017
Total fat (kg)	13.5±3.0	16.8±6.4	0.065
Percentage arm fat (%)	20.5±8.5	24.4±8.7	0.186
Percentage leg fat (%)	28.1±5.6	30.0±7.6	0.421
Percentage trunk fat (%)	24.5±6.4	29.3±7.8	0.058
Percentage body fat (%)	24.6±5.6	28.0±7.3	0.136
Trunk/leg fat ratio	1.10±0.23	1.33±0.23	0.006
Glucose: fasting (mg/dL)	84±5	84±4	0.708
$30 \min (mg/dL)$	113±12	$100{\pm}14$	0.007
$60 \min(mg/dL)$	91±19	78±16	0.027
$120 \min (mg/dL)$	76±12	73±10	0.507
Insulin: fasting (µÚ/mL)	3.1±1.1	4.3±2.5	0.078
$30 \min (\mu U/mL)$	29.1±15.7	36.8±19.3	0.206
$60 \min(\mu U/mL)$	28.1±20.0	27.2±13.1	0.871
120 min (µU/mL)	11.3±7.8	13.6±8.2	0.399
HbA1c (%)	4.7±0.2	4.8±0.2	0.232
HOMA-IR	0.63±0.21	0.89±0.54	0.077
log HOMA-IR	-0.22±0.14	-0.13±0.28	0.244
Glucose-AUC (mg/dL. 2 h)	184±16	166±20	0.007
Insulin-AUC (µU/mL. 2 h)	42.0±25.1	46.7±22.0	0.566
Cholesterol (mg/dL)	168±18	188±22	0.006
TG: fasting (mg/dL)	41±10	68±16	0.000
$30 \min (mg/dL)$	43±11	78±17	0.000
$60 \min(mg/dL)$	51±12	94±16	0.000
120 min (mg/dL)	56±14	102±22	0.000
TG-AUC $(mg/dL. 2h)$	98±22	177±27	0.000
HDL cholesterol (mg/dL)	75±13	73±10	0.574
Apolipoprotein A1 (mg/dL)	158±14	160±15	0.689
Apolipoprotein B (mg/dL)	59±9	73±14	0.001
PAI-1 (ng/mL)	29±27	30±11	0.984
$TNF-\alpha$ (pg/mL)	0.6±0.1	0.7±0.3	0.211
Leptin (ng/mL)	5.1±2.2	8.8±5.1	0.010
Adiponectin (µg/mL)	12.4±4.9	10.1 ± 3.8	0.123
Leptin/adiponectin ratio	$0.44{\pm}0.20$	$0.98{\pm}0.64$	0.003
IL-6 (pg/mL)	0.7±0.4	0.6±0.2	0.244
hsCRP (μ g/dL)	28±49	55±118	0.383
log hsCRP	1.09±0.49	1.33±0.52	0.171

Table 4. Anthropometric, biochemical and postprandial variables of young women stratified into 2 groups according to the median area under the curve of triglycerides (TG-AUC)

Abbreviations are the same as in Table 1. Mean±SD.

Table 5. Associations	(correlation coefficients) of trunk/leg fat ratio	, trunk and leg fat with	serum triglyceride levels
during meal tests				

	Trunk/leg fat ratio	Trunk fat		Leg fat	
	Simple	Simple	Leg fat adjusted	Simple	Trunk fat adjusted
TG 0 min	0.450**	0.262	0.511**	0.049	-0.487**
TG 30 min	0.524**	0.299	0.585^{**}	0.044	-0.537**
TG 60 min	0.607^{**}	0.399^{*}	0.665^{**}	0.097	-0.571**
TG 120 min	0.413*	0.211	0.543**	-0.007	-0.492**
TG-AUC	0.549**	0.327	0.630**	0.053	-0.563**

p*<0.05; *p*<0.01.

Abbreviations are the same as in Table 1.

apolipoprotein B metabolism have been reported to occur in the early phase of insulin resistance with abdominal obesity.²¹ Although insulin resistant women with abdominal obesity²¹ were normoglycemic and fasting normotriglyceridemic, waist and HOMA-IR averaged 125 cm and 4.1, respectively. In contrast, waist and HOMA-IR averaged 75.4 cm and 0.89, respectively, in our young women with higher high TG-AUC. These findings sug-

gest that modest increase in trunk fat may be associated with decrease in insulin sensitivity and elevated postprandial lipemia and apoB.

Association of post-meal TG with MSIS in our young women may be compatible with a study showing that not only resistance to insulin-mediated glucose disposal (as quantified by determining the steady-state plasma glucose concentration during the insulin suppression test) but the meal-induced insulin response was an independent predictor of degree of postprandial lipemia in healthy, nondiabetic subjects.²² There are ample data showing that the plasma insulin response is highly correlated with degree of insulin resistance.^{23,24}

Long-term endurance training has been shown to result in significant improvements in plasma lipids and lipoprotein profiles, including fasting TG, of sedentary, overweight subjects with mild to moderate dyslipidemia.²⁵ In the present study, however, long-term endurance training had no effect on triglyceridemia. This may be because of low fasting and postprandial triglyceridemia in a sedentary group, non-athletes: their mean TG increased from 54 at baseline to 78 mg/dL at 120 min after ingestion of test meals.

This study has several strengths, including a homogeneous study population with scarce confounding factors, and accurate and reliable measures of body composition by DXA. The main limitations of our study are small sample size and small amount of TG loaded: standardized meals contained only 17 g of TG. Although this amount when expressed by energy percentage is higher (33%) than usual Japanese breakfast (20-25%), this may be too small to evaluate postprandial TG response and 2 h of follow-up period may be short. However, mean increase in TG from 55 to 79 mg/dL by 24 mg/dL in our young women appeared to be comparable to TG increase by 31 mg/dL over baseline at 2 h after 15-g fat meals in young healthy men.²⁶ DXA does not allow separate quantification of visceral fat and subcutaneous fat in the trunk. However, the association of trunk fat with HOMA-IR was comparable to that of visceral adipose tissue accumulation by computed tomography.²⁷ The cross-sectional design of the present study complicates the drawing of causal inferences, and a single measurement of biochemical variables may be susceptible to short-term variation, which would bias the results toward the null. We used several surrogates in the present study, which may be less accurate.

Conclusion

Trunk/leg fat ratio, a marker of central fat accumulation, is a significant predictor of postprandial lipemia even in young, lean and insulin-sensitive women independently of adiposity, insulin resistance, serum adiponectin and inflammation, suggesting a modifiable pathway to postprandial hypertriglyceridemia, a risk factor for coronary heart disease among Japanese men and women whose non-fasting TG averaged 145 and 136 mg/d/L, respective-ly.²⁸ Recently, baseline and change in visceral adipose tissue have been reported to be independent predictors for future development of atherogenic dyslipidemia in Japanese Americans.²⁹

ACKNOWLEDGEMENTS

We are indebted to all the participants for their dedicated and conscientious collaboration.

AUTHOR DISCLOSURES

The authors declare no conflict of interest. No competing financial interests exist.

REFERENCES

- Goldberg IJ, Eckel RH, McPherson R. Triglycerides and heart disease: still a hypothesis? Arterioscler Thromb Vasc Biol. 2011;31:1716-25. doi: 10.1161/ATVBAHA.111.226 100.
- Nordestgaard BG. Triglyceride-rich lipoproteins and atherosclerotic cardiovascular disease: new insights from epidemiology, genetics, and biology. Circ Res. 2016;118: 547-63. doi: 10.1161/CIRCRESAHA.115.306249.
- 3. Zilversmit DB. Atherogenesis: a postprandial phenomenon. Circulation. 1979;60:473-85.
- SnidermanAD. Postprandial hypertriglyceridemia(s): time to enlarge our pathophysiologic perspective. Eur J Clin Invest. 2000;30:935-7.
- Patsch JR, Miesenbock G, Hopferwieser T, Mühlberger V, Knapp E, Dunn JK, Gotto AM Jr, Patsch W. Relation of triglyceride metabolism and coronary artery disease. Studies in the postprandial state. Arterioscler Thromb. 1992;12: 1336-45.
- Pirillo A, Norata GD, Catapano AI. Postprandial lipemia as a cardiometabolic risk factor. Curr Med Res Opin. 2014;30: 1489-503. doi: 10.1185/03007995.2014.909394.
- Borén J, Matikainen N, Adiels M, Taskinen MR. Postprandial hypertriglyceridemia as a coronary risk factor. Clin Chim Acta. 2014;431:131-42. doi: 10.1016/j.cca.2014. 01.015.
- Lewis GF, O'Meara NM, Soltys PA, Blackman JD, Iverius PH, Druetzler AF, Getz GS, Polonsky KS. Postprandial lipoprotein metabolism in normal and obese subjects: comparison after the vitamin A fat-loading test. J Clin Endocrinol Metab. 1990;71:1041-50.
- Couillard C, Bergeron N, Prud'homme D, Bergeron J, Tremblay A, Bouchard C, Mauriège P, Després JP. Gender difference in postprandial lipemia: importance of visceral adipose tissue accumulation. Arterioscler Thromb Vasc Biol. 1999;19:2448-55.
- Halkes CJ, Castro Cabezas M, van Wijk JP, Erkelens DW. Gender differences in diurnal triglyceridemia in lean and overweight subjects. Int J Obes Relat Metab Disord. 2001; 25:1767-74.
- JangY, Kim OY, Ryu HJ, Kim JY, Song SH, Ordovas JM, Lee JH. Visceral fat accumulation determines postprandial lipemic response, lipid peroxidation, DNA damage, and endothelial dysfunction in nonobese Korean men. J Lipid Res. 2003;44:2356-64.
- Blackburn P, Lamarche B, Couillard C, Pascot A, Tremblay A, Bergeron J, Lemieux I, Després JP. Contribution of visceral adiposity to the exaggerated postprandial lipemia of men with impaired glucose tolerance. Diabetes Care. 2003; 26:3303-09.
- Alipour A, Elte JW, van Zaanen HC, Rietveld AP, Castro Cabezas M. Novel aspects of postprandial lipemia in relation to atherosclerosis. Atheroscler Suppl. 2008;9:39-44. doi: 10.1016/j. atherosclerosissup.2008.05.007.
- 14. Tanaka S, Wu B, Honda M, Suzuki K, Yoshino G, Fukuo K, Kazumi T. Associations of lower-body fat mass with favorable profile of lipoproteins and adipokines in healthy, slim women in early adulthood. J Atheroscler Thromb. 2011; 18:365-72. doi: 10.5551/jat.7229.

- Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. Diabetologia. 1985;28:412-9.
- 16. Finucane FM, Luan J, Wareham NJ, Sharp SJ, O'Rahilly S, Balkau B et al. Correlation of the leptin: adiponectin ratio with measures of insulin resistance in non-diabetic individuals. Diabetologia. 2009;52:2345-49. doi: 10.1007/ s00125-009-1508-3.
- Tanaka M, Yoshida T, Bin W, Fukuo K, Kazumi T. FTO, abdominal adiposity, fasting hyperglycemia associated with elevated HbA1c in Japanese middle-aged women. J Atheroscler Thromb. 2012;19:633-42. doi: 10.5551/jat.119 40.
- 18. Wu H, Qi Q, Yu Z, Sun Q, Wang J, Franco OH et al. Independent and opposite associations of trunk and leg fat depots with adipokines, inflammatory markers, and metabolic syndrome in middle-aged and older Chinese men and women. J Clin Endocrinol Metab. 2010;95:4389-98. doi: 10.1210/jc.2010-0181.
- Sakai Y, Ito H, Egami Y, Ohoto N, Hijii C, Yanagawa M, Satoh S, Jingu S. Favourable association of leg fat with cardiovascular risk factors. J Intern Med. 2005;257:194-200. doi: 10.1111/j.1365-2796.2004.01432.x
- 20. Lim U, Turner SD, Franke AA, Cooney RV, Wilkens LR, Ernst T et al. Predicting total, abdominal, visceral and hepatic adiposity with circulating biomarkers in Caucasian and Japanese American women. PLoS One. 2012;7:e43502. doi: 10.1371/journal.pone.00435 02.
- 21. Pont F, Duvillard L, Florentin E, Gambert P, Vergès B. Early kinetic abnormalities of apoB-containing lipoproteins in insulin-resistant women with abdominal obesity. Arterioscler Thromb Vasc Biol. 2002;22:1726-32. doi: 10. 1161/01.ATV.0000032134.92180.41.

- 22. Jeppesen J, Hollenbeck CB, Zhou MY, Coulston AM, Jones C, Chen YD, Reaven GM. Relation between insulin resistance, hyperinsulinemia, postheparin plasma lipoprotein lipase activity, and postprandial lipemia. Arterioscler Thromb Vasc Biol. 1995;15:320-4.
- Hollenbeck C, Reaven GM. Variations in insulin-stimulated glucose uptake in healthy individuals with normal glucose tolerance. J Clin Endocrinol Metab. 1987;64:1169-73.
- Reaven GM, Brand RJ, Chen Y-DI, Mathur AK, Goldfine I. Insulin resistance and insulin secretion are determinants of oral glucose tolerance in normal individuals. Diabetes. 1993; 42:1324-32.
- 25. Kraus WE, Houmard JA, Duscha BD, Knetzger KJ, Wharton MB, McCartney JS et al. Effects of the amount and intensity of exercise on plasma lipoproteins. N Engl J Med. 2002:347:1483-92.
- 26. Dubois C, Beaumier G, Juhel C, Armand M, Portugal H, Pauli AM, Borel P, Latgé C, Lairon D. Effects of graded amounts (0-50 g) of dietary fat on postprandial lipemia and lipoproteins in normolipidemic adults. Am J Clin Nutr. 1998; 67:31-8.
- 27. Müller MJ, Lagerpusch M, Enderle J, Schautz B, Heller M, Bosy-Westphal A. Beyond the body mass index: tracking body composition in the pathogenesis of obesity and the metabolic syndrome. Obes Rev. 2012;13(Suppl 2):6-13. doi: 10.1111/j.1467-789X.2012.01 033.x.
- 28. Iso H, Naito Y, Sato S, Kitamura A, Okamura T, Sankai T, Shimamoto T, Iida M, Komachi Y. Serum triglycerides and risk of coronary heart disease among Japanese men and women. Am J Epidemiol. 2001;153:490-9.
- 29. Hwang YC, Fujimoto WY, Hayashi T, Kahn SE, Leonetti DL, Boyko EJ. Increased visceral adipose tissue is an independent predictor for future development of atherogenic dyslipidemia. J Clin Endocrinol Metab. 2016;101:678-5. doi: 10.1210/jc.2015-3246.